NEW POLY (ETHYLENE GLYCOL) SUPPORTS FOR THE LIQUID-PHASE SYNTHESIS OF

PROTECTED PEPTIDE HYDRAZIDES

Roberto Colombo

St. Raphael Hospital Research Department, School of Medicine, University of Milan, via Olgettina 60, I-20132 Milan, Italy.

Summary. The liquid-phase synthesis of protected peptide hydrazides can be suitably per formed on modified poly(ethylene glycol) supports containing benzyl-, p-benzyloxybenzyl-, and *tert*-alkyl-oxycarbonylhydrazide anchoring groups.

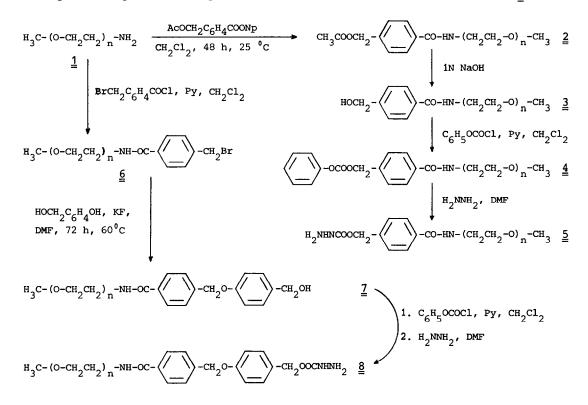
Protected peptide hydrazides¹ are useful precursors for the synthesis of the long peptide chains in that reaction with aqueous nitrous acid or alkyl nitrite in organic solvents gives the corresponding azides, which are suitable for seg ment condensation. The liquid-phase method² for peptide synthesis offers a rapid procedure, which combines strategic elements of both classical and solid-phase technique, for assembling small and medium-sized peptide chains on a poly meric soluble support. Consequently, protected peptide hydrazides may be ob tained by either hydrazinolysis of peptide chains linked to poly(ethylene glycol) supports through an ester bond^{1,3} or direct coupling of released protected peptide acids with hydrazine⁴. Though both of these procedures are attractive, they are not without some undesirable limitations and complications, such as low yields or modifications of Asp and Glu side chains protected as benzyl-type esters, and a more generally applicable method is desirable.

Recently a number of modified poly(ethylene glycol) supports were described ^{3,5}, which can be used for the liquid-phase synthesis of free and protected pep tides and of peptide amides⁵, and are cleavable from the finished product by acidolysis, hydrogenolysis, and photolysis; I wish now to report the preparation and use of a new type of modified soluble polymers, bearing an oxycarbonylhydra zyde functional group and designed for the direct liquid-phase synthesis of peptide hydrazides.

Poly(ethylene glycol) monomethyl ether (M_r = 5000) was converted to amino-poly (ethylene glycol) monomethyl ether <u>1</u> according to the procedure of Mutter⁶, and <u>1</u> was allowed to react with the p-nitrophenyl ester of p-acetyloxymethyl-

4129

benzoic acid (prepared in the usual manner²⁰); the product, $\underline{2}$, was deacetylated with 1 N NaOH to yield $\underline{3}$, which was acylated with phenyl chloroformate, and the phenyl carbonate ($\underline{4}$) obtained was then hydrazinolyzed to give the benzyloxyca<u>r</u> bonylhydrazide polymer $\underline{5}$. The amino-polymer $\underline{1}$ was also converted to its p-bro<u>o</u> momethylbenzoyl derivative $\underline{6}$ by treatment with p-bromethylbenzoyl chloride⁵, and the substituted benzyl bromide $\underline{6}$ was allowed to react with a large excess (5-8 equiv.) of 4-hydroxymethylphenol in the presence of KF⁷ to give $\underline{7}$; the reaction of the alkoxybenzyl alcohol $\underline{7}$ with phenyl chloroformate first and then with hydrazine yielded the p-alkoxybenzyloxycarbonylhydrazide polymer $\underline{8}$.



The monofunctional poly(ethylene glycol) bearing a tertiary alcohol group $\underline{9}$, prepared as described by Anzinger *et al.*⁸, was converted to the *tert*-alkyloxycarbonylhydrazide polymer $\underline{10}$ by treatment with phenyl chloroformate followed by hydrazinolysis. Polymers $\underline{5}$, $\underline{8}$, and $\underline{10}$ were repeatedly crystallized from CH₂Cl₂ and DMF solutions by slow addition of anhydrous diethyl ether untill chromatographically homogeneous. Hydrazide contents were determined by nitrogen microanalysis.

The modified poly(ethylene glycol) supports $\underline{5}$, $\underline{8}$, and $\underline{10}$ were employed for as sembling a number of protected tri-, tetra-, and pentapeptide hydrazides (*Table*

PRODUCT ^a		CLEAVAGE ^b : Agent Yield%		overall yield %	ு மை (oC)	$\left[\alpha\right]_{\rm D}^{25}$, deg		Ref.
······································		Agent	Yield %					
Boc-Ala-Ala-Ala-NHNH ₂	5	н2	96	53	247-249	-18.1±0.4	(c=1, DMF)	(đ)
Boc-Leu-Leu-Gly-NHNH ₂	5	^н 2	91	48	oil ^e			(f)
Boc-Gln-Ala-Ala-Thr- Gly-NHNH ₂	5	^н 2	95	35	207-209	-15.2±0.5	(c=0.5, DMF)	(g)
Boc-Ala-Glu (OBu ^t) -Lys (N ^{ε} -Boc) -Lys (N ^{ε} Boc) - NHNH ₂	5	^H 2	90	27	167-168	-23.8±0.3	(c=1, CH ₃ OH)	(h)
Fmoc-Gly-Ala-Val-Leu- NHNH ₂	8	TFA	92	43	218-222	-26.5±0.5	(c=1, DMF)	(i)
Fmoc-Gly-Phe-Phe-Tyr (Bzl)-Thr(Bzl)-NHNH ₂	8	TFA	85	41	190-193	- 1.2±0.2	(c=1, DMF)	(j)
Z-Gly-Leu-Phe-NHNH ₂	8≝	TFA	93	49	193-194	-29.7±0.3	(c=0.5, DMF)	(k)
$z-val-Lys(N^{\epsilon}-z)-Pro-Gly-NHNH_2$	₿	TFA	89	37	202-203	-44.1±0.5	(c=1, CH ₃ OH)	(1)
	<u>10</u>	TFA	9 5	35	203-204	-43.0±0.4	(c=1, CH ₃ OH)	(1)
Z-Gly-Phe-Phe-Tyr (Bzl)-Thr(Bzl)-NHNH 2	툍	TFA	87	31	226-227	- 2.1±0.5	(c=1, DMF)	(m)
	<u>10</u>	TFA	94	30	225-227	- 2.9±0.4	(c=1, DMF)	(m)

Table 1. Protected Peptide Hydrazides prepared by Liquid-Phase Method.

^a Satisfactory elemental and amino acids analyses have been obtained for all products. ^b The reported cleavage yields are based on quantitative amino acid analysis of the peptide remaining on the polymer after removal of the cleaved peptide hydrazide. ^c Based on starting protected aminoacyl-polymer. ^d Ref. 9 : mp 249-250, OR -17.3±0.3. ^e T.1.c. on silica gel plates (Merck, F-254) gave one chlorine positive, picryl-positive spot, R_f 0.58 (n-BuOH/AcOH/H₂O 5:1:4); microanalysis was performed on the tripeptide hydrazide obtained after removal of the Boc N^Q-protecting group by 50% TFA in CH₂Cl₂. ^f Ref. 10. ^g Ref. 11: mp 210-212, OR -16.2 (c=0.4, DMF). ^h Ref. 12: mp 165-167, OR -25.0 (c=1.1, CH₃OH). ⁱ Ref. 13: mp 220-225, OR -24.25. ^j Ref. 13: mp 196-198, OR -0.40. ^k Ref. 14: mp 191-192, OR -28.3. ¹ Ref. 15: mp 200-201, OR -47.7. ^m Ref. 16: mp 215-218, OR -3.94.

1), which after cleavage from the polymers and crystallization from a suitable solvent (CH₃OH, C₂H₅OH, or DMF by addition of water or diethyl ether) were obtained in satisfactory yields and homogeneous form. At the start N^{α}-protected amino acids were condensed with the hydrazide supports by means of DCC in the presence of 1 equiv. of HOBt¹⁷, and the stepwise syntheses were then performed by the DCC/HOBt procedure according to the general techniques of liquid-phase method².

The anchoring bond between the peptide chain and polymer 5 was cleaved, at the end of the synthesis, by catalytic hydrogenolysis (5% Pd on BaSO₄, 40 psi,

room temperature, 18-22 h), thus leaving tert-butyl based protecting groups un affected. Bonds formed by supports <u>8</u> and <u>10</u>, which are structurally similar to those devised by Wang and Merrifield¹⁸ and by Wang¹⁹ for the solid-phase syn thesis of peptide hydrazides, are acid-labile and were selectively cleaved by 50% TFA in CH₂Cl₂ (30-60 min, 25 °C); under these conditions the benzyl based protecting groups are normally stable as well as the Fmoc group, thus allowing the preparation of fully protected peptide hydrazides. Syntheses on support <u>5</u> were performed employing, as N^{α} temporary protection, the Boc group (if no side chain protecting groups were present) or the Bpoc group (if tert-butyl based protecting groups were introduced during the chain assembly); the last amino acid was coupled as its N^{α}-Boc derivative. The synthesis on support <u>8</u> is com patible with Bpoc and Fmoc N^{α} protection, while polymer <u>10</u> can be used with a strategy of synthesis based on the use of N^{α}- Bpoc, Fmoc, or Z groups.

REFERENCES AND NOTES

Abbreviations: Boc, tert-butyloxycarbonyl; Bpoc, 2-(4-biphenylyl)propyl(2)oxycarbonyl; Bu^C, tert-butyl; Bzl, benzyl; DCC, dicyclohexylcarbodiimide; Fmoc, 9-fluorenylmethyloxycarbonyl; HOBt, 1-hydroxybenzotriazole; ONp, 4-nitrophenyl ester; OR, optical rotation; Py, pyridine; TFA, trifluoroacetic acid; Z, benzyloxycarbonyl.

- J. Meienhofer, in: The Peptides (E. Gross & J. Meienhofer eds), vol. 1, pp. 198-237, Academic Press, New York (1979).
- M. Mutter and E. Bayer, in: The Peptides (E. Gross & J. Meienhofer eds), vol. 2, pp. 285-332, Academic Press, New York (1980).
- 3. F.-S. Tjoeng, E.K. Tong, and R.S. Hodges, J. Org. Chem. 43 , 4190 (1978).
- 4. Protected peptide hydrazides may be prepared from the corresponding acids by DCC coupling in the presence of additives (HOBt, N-hydroxysuccinimide) for racemization suppression; see Ref. 1 and references cited therein.
- 5. V.N.R. Pillai, M. Mutter, E. Bayer, and I. Gatfield, J. Org. Chem. 45, 5364 (1980).
- 6. M. Mutter, Tetrahedron Lett. 2839 (1978).
- 7. J.P. Tam, R.D. Dimarchi, and R.B. Merrifield, Int. J. Peptide Protein Res. 16, 412 (1980).
- 8. H. Anzinger, M. Mutter, and E.Bayer, Angew. Chem. 91, 747 (1979).
- 9. L. Moroder, F. Marchiori, R. Rocchi, A. Fontana, and E. Scoffone, J. Am. Chem. Soc. <u>91</u>, 3921 (1969).
- 10. G.R. Pettit and W.R. Jones, J. Org. Chem. 36, 870 (1971).
- 11. E. Schaich and F. Schneider, Hoppe-Seyler's Z. Physiol. Chem. 355, 952 (1974).
- 12. H. Yajima, K. Kawasaki, H. Minami, H. Kawatani, N. Mizokami, Y. Kiso, and F. Tamura, Chem. Pharm. Bull. 18, 1394 (1970).
- 13. S.S. Wang, J. Org. Chem. 40, 1235 (1975).
- 14. K. Bláha, Collect. Czech. Chem. Comm. 37, 1861 (1972).
- 15. E. Schaich and F. Schneider, Hoppe-Seyler's Z. Physiol. Chem. 355, 939 (1974).
- 16. J.K. Chang, M. Shimizu, and S.S. Wang, J. Org. Chem. 41, 3255 (1976).
- 17. W. König and R. Geiger, Chem. Ber. 103 , 788 (1970).
- 18. S.S. Wang and R.B. Merrifield, J. Am. Chem. Soc. 91, 6488 (1969).
- 19. S.S. Wang, J. Am. Chem. Soc. <u>95</u>, 1328 (1973).
- 20. M. Bodanszky and V. du Vigneaud, J. Am. Chem. Soc. 81, 5688 (1959).

(Received in UK 6 July 1981)